

The maximum degree of inactivation so far obtained has been $80 \pm 5\%$, as measured in the standard assay.³ To determine whether this represents a mixture of 20% of active chymotrypsin with 80% of totally inactive enzyme or an enzyme possessing 20% activity, samples of the inactivated enzyme were allowed to react with diisopropylfluorophosphate. Since it is known that during the complete inactivation of chymotrypsin by diisopropyl fluorophosphate,⁷ one mole of diisopropyl phosphate per mole of enzyme⁷ is bound to the serine hydroxyl group at the active site,⁸ the phosphorus content of inhibited chymotrypsin, allowed to react with diisopropylfluorophosphate until completely inactive, is a measure of the proportion of enzyme molecules capable of reaction at the active site.⁹ The phosphorylated proteins, which had less than 0.25% activity, contained¹⁰ one mole of phosphorus per mole of enzyme (Table I).

Amino acid analyses¹¹ of chymotrypsin and of the irreversibly inactivated chymotrypsin show that one methionine residue in the latter is destroyed, and that the products (after hydrolysis) from it are the same as those obtained by Gundlach, Stein, and Moore (ref. 12, fig. 2c) with ribonuclease inactivated by iodoacetic acid at pH 2.8. In addition, roughly one mole of α -aminoisobutyric acid is found in the hydrolysate (Table II). No other change in amino acid composition has been detected.

TABLE II
METHIONINE, ITS DECOMPOSITION PRODUCTS AND α -AMINOISOBUTYRIC ACID IN HYDROLYSATES OF MODIFIED CHYMOTRYPSIN

Amino acid	Modified CT	CT	DIP-CT ^a
Methionine	1.19 ^b	2.08 ^b	2.10
S-Carboxymethylhomocysteine	0.5	...	ca. 0.01
Homoserine	0.1	...	0.00
Homoserine lactone	0.3	...	0.00
α -Aminoisobutyric acid	ca. 1.0 ^c	...	trace

^a The DIP (diisopropylphosphoryl)-CT, which had 0.25% activity, was allowed to react with 25 moles of I for 6 hr. ^b Average of three determinations. ^c This amino acid was difficult to determine with accuracy since it gives a low ninhydrin color yield and falls on one side of the cystine peak in chromatograms.¹¹

We conclude that the activated bromine atom of the acyl enzyme alkylates a methionine residue in the vicinity of the active site, either while attached

(7) E. F. Jansen, M.-D. F. Nutting and A. K. Balls, *J. Biol. Chem.*, **179**, 201 (1949); A. K. Balls and E. F. Jansen, *Advances in Enzymol.*, **13**, 321 (1952).

(8) J. A. Cohen, R. A. Oosterbaan, H. S. Jansz and F. Berends, *J. Cell. Comp. Physiol.*, **54**, Suppl. 1, 231 (1959), and refs. cited therein.

(9) This method is analogous to the "all or none" assays of (a) W. J. Ray, Jr., J. J. Ruscica and D. E. Koshland, Jr., *J. Am. Chem. Soc.*, **82**, 4739 (1960), and (b) W. J. Ray, Jr. and D. E. Koshland, Jr., Abstracts of Papers, American Chemical Society Meeting, Washington, D. C., March 21-24, 1962, p. 41-C.

(10) "Official and Tentative Methods of Analysis of the Association of Official Agricultural Chemists," 6th Ed., A.O.A.C., Washington, D. C., 1945, p. 127.

(11) D. H. Spackman, W. H. Stein and S. Moore, *Anal. Chem.*, **30**, 1190 (1958).

(12) (a) H. G. Gundlach, W. H. Stein and S. Moore, *J. Biol. Chem.*, **234**, 1754 (1959); (b) H. G. Gundlach, S. Moore and W. H. Stein, *ibid.*, **234**, 1761 (1959).

to the serine or shortly after hydrolysis of the acyl-serine bond. Since the liberated serine hydroxyl group is capable of reaction with diisopropylfluorophosphate, the active site is still capable of functioning, but much less efficient than it was before modification of the methionine residue near it. The inefficiency of the modified enzyme is attributable to a considerable increase (about 11-fold) in the Michaelis constant over that of native chymotrypsin.¹³

Our work corroborates the finding of Ray, *et al.*,¹⁴ that destruction of methionine leads to the inactivation of chymotrypsin. It is unlikely that the sulfide group of the critical methionine behaves as a neighboring nucleophile at the active site of chymotrypsin (though this is entirely possible in the case of phosphoglucomutase¹⁴), since this activity should be abolished upon alkylation, which gives a ternary sulfonium salt,^{12,15} or upon subsequent peptide cleavage.^{15,16} The increase in the Michaelis constant indicates that modification of the critical methionine residue results in decreased affinity for substrate.

This general method for the modification of enzymes, as well as the complementary method of Baker, *et al.*,¹⁷ should prove to be a good tool for the "mapping" of enzyme active sites.

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(13) Procedure of L. W. Cunningham, Jr., *ibid.*, **207**, 443 (1954).

(14) W. J. Ray, Jr., H. G. Latham, Jr., M. Katsoulis and D. E. Koshland, Jr., *J. Am. Chem. Soc.*, **82**, 4743 (1960).

(15) W. B. Lawson, E. Gross, C. M. Foltz and B. Witkop, *ibid.*, **83**, 1509 (1961); E. Gross and B. Witkop, *ibid.*, **83**, 1510 (1961).

(16) Subsequent peptide cleavage at the carboxyl group of the methionine is likely to be slight under the conditions used (*cf. ref. 15*). This point, as well as the possibility of cleavage upon heating, is under investigation.

(17) B. R. Baker, W. W. Lee, E. Tong and L. O. Ross, *J. Am. Chem. Soc.*, **83**, 3713 (1961); *cf. also* G. Schoellmann and E. Shaw, *Biochem. Biophys. Res. Comm.*, **7**, 36 (1962); *Fed. Proc.*, **21**, 232 (1962).

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A STEREOSPECIFIC TOTAL SYNTHESIS OF 18-SUBSTITUTED STEROIDS. APPLICATION TO THE SYNTHESIS OF *dl*-CONESSINE

Sir:

This communication reports a total synthesis of 18-substituted steroids in which the B-C-D system is constructed following the lines laid in our previous steroid total synthesis,¹ but control of the C₁₀ stereochemistry is achieved by the use of an 8-*iso* (*cis* B/C) structure which finally is inverted to the 8-normal (β) configuration. We illustrate the synthesis by building up the steroid alkaloid conessine,² but it will be obvious that a variety of 18-substituted 20-ketosteroids would be of considerably simpler access, *e.g.*, *via* IV.

(1) G. Stork, H. J. E. Loewenthal and P. C. Mukharji, *J. Am. Chem. Soc.*, **78**, 501 (1956).

(2) D. H. R. Barton and L. R. Morgan, *J. Chem. Soc.*, 622 (1962). have accomplished a partial synthesis of natural conessine from Δ^4 -pregnene-3 β ,20 β -diol.

anhydride-sodium acetate) into the corresponding enol lactone m.p. 196–197° (C, 73.55; H, 8.45) which readily gave the pentacyclic enone XIII, m.p. 142–143° (150°) (C, 77.45; H, 9.20 infrared identical with XIII *ex* natural conessine) on treatment with ethereal methylmagnesium iodide. Formation of the N,N-dimethylenamine with methanolic dimethylamine and *p*-toluenesulfonic acid was followed by its reduction with sodium borohydride-diglyme in a mixture of aqueous ethanol, acetic acid and sodium acetate,⁴ and the resulting 3β-dimethylamino compound was then deacetylated (calcium in liquid ammonia) and methylated by the usual formic acid-formaldehyde procedure to give *dl*-conessine (XIV), m.p. 127.5–128.5° (Kofler). The identity of the infrared spectrum (CCl₄) with that of an authentic sample of the natural material establishes the suggested stereochemical course of the various synthetic steps.⁵

(4) This procedure is adapted from that first described by W. S. Johnson, *et al.* (*Tetrahedron Letters*, No. 2, 72 (1961)).

(5) We thank the National Science Foundation and the National Institutes of Health for their support of this work.

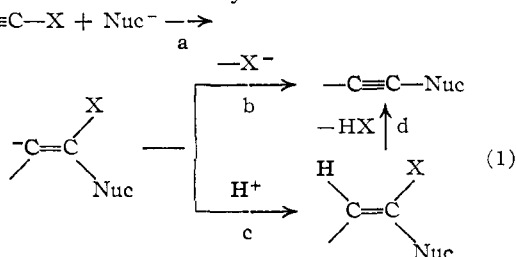
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NUCLEOPHILIC SUBSTITUTION AT AN ACETYLENIC CARBON

Sir:

Analogs of the ordinary displacement reaction are now known in the benzene¹ and ethylene series.² Based on several negative reports, it is generally considered that haloalkynes are inert to nucleophilic displacement.



Otherwise, the formation of $\text{—C}\equiv\text{C—Nuc}$ in the presence of a proton donor often has been the over-all result of the now familiar addition elimination sequence, a–c–d in eq. 1;^{3,5} indeed, the presence of a proton donor often renders the substitution mechanism uncertain.⁶ Ott and

(1) J. F. Bunnett, *Quart. Revs.*, **12**, 1 (1958).

(2) S. I. Miller and P. K. Yonan, *J. Am. Chem. Soc.*, **79**, 5931 (1957).

(3) (a) J. A. Nieuwland and R. R. Vogt, "The Chemistry of Acetylene," Reinhold Publishing Corp., New York, N. Y., 1945, p. 71; M. J. Murray, *J. Am. Chem. Soc.*, **60**, 2662 (1938); (b) W. E. Truce, H. E. Hill, and M. M. Boudakian, *ibid.*, **78**, 2760 (1956); (c) R. Truchet, *Ann. chim.*, (10) **16**, 309 (1931); V. Grignard and H. Perrichon, *ibid.*, (10) **5**, 5 (1926); J. Loevenich, J. Losen, and A. Dierichs, *Ber.*, **60**, 950 (1927); J. U. Nef, *Ann.*, **308**, 264 (1899); V. Wolf and E. Kowitz, *ibid.*, **638**, 33 (1960); V. Wolf and W. Block, *ibid.*, **637**, 119 (1960); J. H. Boyer, C. H. Mack, N. Goebel, and L. R. Morgan, Jr., *J. Org. Chem.*, **23**, 1051 (1958).

(4) E. Ott and G. Dittus, *Ber.*, **76**, 80 (1943).

(5) S. I. Miller, *J. Org. Chem.*, **26**, 2619 (1961); W. E. Truce, M. M. Boudakian, R. F. Heine, and R. J. McManis, *J. Am. Chem. Soc.*, **78**, 2743 (1956).

(6) (a) H. J. Boonstra and J. F. Arens, *Recueil*, **79**, 866 (1960); J. R. Nooi and J. F. Arens, *ibid.*, **80**, 244 (1961); (b) G. Luciani and F.

Dittus do describe what appears to be close to an authentic displacement, *e.g.*, dichloroethyne with diethyl sodioethylmalonate in ether.^{4,7}

We have observed that sodium thiolates react with haloalkynes in dimethylformamide (DMF) according to a–b in eq. 1. Thus we have obtained 1-phenyl-2-phenylthioethyne, 1-phenyl-2-pentachlorophenylthioethyne, and 1-phenyl-2-*p*-tert-butylphenylthioethyne. Of course, all of the products mentioned in this paper have given satisfactory elemental analyses. Of these, only the 1-phenyl-2-phenylthioethyne appears to be known; its properties and those of the derived sulfone check with those previously reported.^{8b} Provided that the parent acetylene is available, the displacement path compares favorably with standard routes to thioethers.⁸

Typical of these preparations was that of 1-phenyl-2-*p*-toluenethioethyne. Sodium toluenethiolate was prepared by treatment of the thiol with sodium in refluxing toluene. The salt was filtered, washed with toluene and vacuum dried. A solution of the sodium toluenethiolate and phenylbromacetylene in DMF was stirred for 1 hr. and then worked up. Chromatography over alumina yielded white crystals, m.p. 45.5–46.5°, in 65% yield (*Anal.* Calcd. for C₁₆H₁₂S: C, 80.31; H, 5.39. Found: C, 80.46; H, 5.50). The sulfone was prepared by oxidation of the thioether with hydrogen peroxide to give a solid, m.p. 80–81° (*Anal.* Calcd. for C₁₆H₁₂SO₂: C, 70.29; H, 4.72. Found: C, 70.0; H, 4.78). The infrared triple bond absorption in the thioether and in the sulfone was at 4.72 μ and 4.74 μ, respectively, in agreement with data for similar compounds.

The proposed mechanism of eq. 1 is of some interest. Nucleophilic displacements at other unsaturated centers, *e.g.*, aromatic,¹ ethylenic,² and carbonyl⁹ presumably have similar intermediates. Such carbanions have been suggested for base-catalyzed alcohol additions to alkynes,¹⁰ and presumably occur in base-catalyzed deuteration of alkenes.¹¹

Arens, *et al.*, use scheme 1 as a matter of course in the reactions of haloalkynes with nucleophiles in liquid ammonia.^{6a} While this mechanism appears to be plausible and useful, it can only be considered as tentative.

The preceding examples taken together with our experience with other nucleophiles, *e.g.*, cyanide and amines, as well as with comparative kinetic data¹² suggest that any preconceived notions of the low nucleophilic reactivity at an acetylenic carbon must be revised. It is true that nucleophilic attack on haloalkynes, even under forcing conditions, has

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(c) F. Moulin, *Helv. Chem. Acta.*, **34**, 2416 (1951).

(7) Barring free radical initiation and catalysis, it seems reasonable to regard the couplings of haloalkynes with Grignard organo-copper or -lithium reagents as related processes involving polar aggregates: H. G. Viehe, *Ber.*, **92**, 3064 (1959); F. Bohmann, P. Herbsu, and H. Gleinig *ibid.*, **94**, 948 (1961).

(8) J. F. Arens, "Advances in Organic Chemistry," R. A. Raphael, E. C. Taylor, and H. Wynberg, editors, Interscience Publishers, Inc., New York, N. Y., 1960, Vol. II, pp. 117 ff.

(9) M. L. Bender, *Chem. Revs.*, **60**, 53 (1960).

(10) S. I. Miller, *J. Am. Chem. Soc.*, **78**, 6091 (1956).

(11) S. I. Miller and W. G. Lee, *ibid.*, **81**, 6313 (1959).

(12) A. K. Kuriakose and S. I. Miller, unpublished results.